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## Transmissibility and Persistence of Oral Polio Vaccine Viruses: Implications for the Global Poliomyelitis Eradication Initiative

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The global poliomyelitis eradication initiative has been a tremendous success, with current evidence suggesting that wild poliovirus will cease to circulate anywhere in the world soon after the year 2000. As the goal of wild poliovirus eradication is approached, concern has been raised about the potential for persistent transmission of oral polio vaccine (OPV) viruses, as these viruses are known to revert toward wild-type neurovirulence. This paper has been extracted from a document prepared for the World Health Organization on the implications of OPV transmissibility for the strategy of stopping OPV vaccination after global eradication of wild polioviruses. The authors review the empirical evidence on OPV transmissibility available from household and community transmission studies and from mass-vaccination experiences. They then consider theoretical measures of transmissibility and persistence for wild and OPV viruses (secondary attack rate, basic reproduction number, and critical populations' size), to assess whether transmissibility of OPV viruses is sufficient to allow persistence of these viruses after cessation of vaccination. The findings indicate that OPV viruses could persist under various plausible circumstances, and that this potential should be a major consideration when planning the cessation of OPV vaccination. *Am J Epidemiol* 1999;150:1001–21.

enterovirus; immunization programs; poliomyelitis; poliovirus vaccine, oral; vaccines

Vaccination against polio has provided one of the triumphs of public health in the 20th century. After their introduction in 1955, these vaccines (first formalininactivated polio vaccine (IPV) and later live oral polio vaccine (OPV)) led to immense reductions in the burden of poliomyelitis disease in country after country. These successes led to a 1988 resolution by the World Health Assembly, committing the World Health Organization to "global eradication of poliomyelitis by the year 2000" (1, p. 2). Progress toward this goal has been impressive, and there is increasing confidence that wild-type polio viruses will cease to circulate in human populations, anywhere in the world, soon after the year 2000.

The global eradication program has emphasized the use of OPV. Among the reasons for this choice are the low cost and the relative logistic ease of administering OPV, as compared with IPV. The mode of action is rapid, through immediate competition with wild viruses in the intestines, in addition to the induction of local (gut) immunity. Furthermore, OPV viruses can be transmitted from vaccinees to their contacts, which results in immunization of some individuals who may be missed by a vaccination program. Although some countries now use IPV or combined IPV-OPV schedules, the vast majority of polio vaccine currently in use is OPV. The problem facing the Polio Eradication Initiative is whether OPV viruses could persist as naturally acquired and transmitted infections in human populations, after cessation of vaccination. While advantageous for vaccination programs, the property of transmissibility, coupled with the fact that OPV

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Abbreviations: AIDS, acquired immunodeficiency syndrome; HIV, human immunodeficiency virus; IPV, formalin-inactivated polio vaccine; OPV, oral polio vaccine;  $R_0$ , basic reproduction number;  $R_n$ , actual (= effective = net) reproduction number; TCID<sub>50</sub>, tissue culture infective dose; VAPP, vaccine-associated paralytic poliomyelitis.

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viruses can, on rare occasions, cause disease, becomes a potential *dis*advantage as we near the global eradication of wild polio. The potential for OPV viruses to persist could threaten the eradication program, given that the overall goal implies the need to rid the world of polioviruses altogether. This concern forms the background to this paper.

Polioviruses. The three serotypes of wild polioviruses differ in the amount of disease for which they are responsible. This has varied among places and over time, reflecting complex epidemiologic dynamics. The relative virulence of the wild viruses (defined as the proportion of paralytic cases among infected individuals) is generally considered to be greatest for type 1 and least for type 2 (2), while their relative transmissibility, measured in various ways discussed below, is thought to be roughly equal. The OPV viruses were derived by serial tissue culture passage attenuation of the wild viruses, which resulted in reduced virulence. Several research groups succeeded in producing strains sufficiently attenuated for vaccine use, but Sabin's strains were the least neurovirulent and are incorporated in all OPV vaccines in use today.

An important feature of polioviruses, and OPV viruses in particular, is their continued genetic variation through mutation and recombination. Molecular studies have identified that attenuation of the Sabin strains is based on just a few base differences for types 2 and 3, with many more for type 1 (3). The attenuating mutations are unstable and, under selection pressures of the human gut, tend to "revert" toward the wild-type sequence within 2-5 weeks of vaccination in 50-100 percent of vaccinees who excrete (4, 5). The issue of reversion is complicated, and the term is used loosely to describe a variety of genetic changes "from" a classic OPV genome "toward" a wild-type genome. Reversions are probably responsible for most of the rare cases of vaccine-associated paralytic poliomyelitis (VAPP), although some cases have been associated with intertypic recombinants (6-8). The full implications of such genetic changes are unclear, but it is not unreasonable to assume that selection pressures under conditions of human-to-human transmission might favor wild-type rather than vaccine-type characteristics. The magnitude and speed of such selection are unknown.

Sources of infection. Polioviruses are enteroviruses, transmitted mainly through the intestinal-fecal-oral route. The viruses are also found in oral secretions (though in lower concentrations) and are thought to be transmitted to some degree via the respiratory-oral route. The relative importance of these routes must depend upon levels of fecal and respiratory hygiene in the population. Overall, wealthier societies have higher levels of fecal hygiene than do poorer societies, as reflected by the relative prevalence of many enteric pathogens. Respiratory hygiene may also be correlated with socioeconomic factors. For example, coughing, sneezing, and spitting behavior probably differ among social groups as do crowding within the home and recirculation of air through ventilation systems. The relation is complex, but it is likely that the *relative* contribution of pharyngeal to fecal-oral transmission of polioviruses differs among societies, and that the *ratio* of pharyngeal as opposed to fecal transmission is positively correlated with socio-economic "development" among societies and with socioeconomic "status" within societies.

As polioviruses generally depend on human (fecaloral) transmission for their persistence in nature, human feces is currently the most important natural source of polioviruses. Once vaccination ceases, the residual vaccine stocks, as well as the feces of vaccinees and of their direct and indirect contacts, will be the most obvious and important sources of OPV viruses, at least in the short term. In the long term, other potential sources must be considered.

Residual vaccine stocks. An obvious potential source of OPV viruses after the cessation of vaccination will be the remaining vaccine vials contained in refrigerators and freezers of health centers, pharmacies, hospitals, and physicians' offices, as well as district, regional, and national vaccine stores throughout the world. Inventory and recovery of these residual stocks will pose a major challenge. This consideration implies that it may be important to stop vaccination everywhere, simultaneously, rather than to allow vaccination to persist in some areas while populations of susceptible individuals accumulate in others.

Persistently infected humans. There is no longterm carrier state for poliovirus in immunocompetent individuals, with the duration of excretion ranging from 1 day to about 4 months (2). However, it has recently been recognized that individuals with various immunologic deficiencies may maintain and excrete polioviruses for many months, even years, providing a potential source for the reintroduction of poliovirus in the future. Immunodeficient individuals most at risk of prolonged enteroviral excretion are those with predominantly antibody defects (e.g., X-linked agammaglobulinemia, immunoglobulin A deficiency, and common variable deficiency), who may shed virus for years in the absence of treatment (9). While cases of VAPP have been reported in individuals with combined immunodeficiencies of B- and T-cell production (10, 11), prolonged excretion has only been documented for those with predominantly antibody defects (12 - 15).

MacCallum (12) reported excretion of OPV-related viruses for up to 32 months by hypogammaglobulinemic individuals, while Hara et al. (13) detected excretion of vaccine-like viruses for at least 34 months by an agammaglobulinemic VAPP case. Recent molecular analyses of poliovirus isolates from a VAPP patient (with common variable immunodeficiency) suggest that OPV viruses may have persisted in the patient for 7 years (15). The pattern of excretion in another 28 hypogammaglobulinemic individuals fed monovalent OPV was the same as that expected for immunocompetent individuals (12), suggesting that chronic infections are rare. Persons with selective immunoglobulin A deficiencies have been shown to have an impaired ability to clear poliovirus infections, although none were observed to excrete OPV viruses after 6 months (14).

Registry data indicate that antibody defects are the most common primary immunodeficiencies. Common variable immunodeficiency prevalence has been estimated at 0.77:100,000 and X-linked agammaglobulinemia at 0.23:100,000 in Australia (16), while immunoglobulin A deficiencies are consistently the most frequent with an estimate of 2:100,000 in Sweden (17). Though the overall incidence of primary immunodeficiencies may not have changed in the last two decades, it is likely that their prevalence in industrialized countries is increasing because of greater recognition and better treatment, enabling increased survival of persons with these conditions. In industrialized countries, the average survival of people treated for antibody defects is greater than 5 years (16) and in some cases can extend beyond 30 years (18). Few data are available on the prevalence and life expectancy associated with primary immunodeficiencies in developing countries. In Sao Paulo, Brazil, individuals with severe combined immunodeficiency had 100 percent mortality over a 15-year period, while those with humoral defects had a mortality rate of just 3 percent, with intravenous immunoglobulin therapy or antibiotic treatment of infections (19). In areas where access to medical care is poor and treatment options are limited, survival rates are likely to be low.

One of the most common immunodeficiencies in developing countries is acquired immunodeficiency syndrome (AIDS), caused by infection with the human immunodeficiency virus (HIV) and primarily affecting T-cells. The implications of HIV infection for the duration of poliovirus or other enterovirus excretion are unknown. To date, only one case of VAPP in an HIVinfected infant has been reported, although no causal relation was established between the HIV infection and VAPP (20). A study of children born to HIV-negative and -positive mothers, in Zaire, found that OPV vaccination induced protective antibody titers in 95 percent of HIV-infected children with no excess risk of vaccine-associated adverse events (21). Although OPV-related adverse events are extremely rare, these data provide no evidence that HIV infection increases the risk of VAPP. If AIDS results in prolonged excretion of poliovirus, these patients may be important sources of OPV in the short term, as AIDS patients survive for a median of 1–2 years with antiretroviral therapy in industrialized countries and for a median of 7–9 months without such treatment in developing country situations (22–24). It is important that more information is obtained regarding this possible long-term source of polioviruses (25).

Nonhuman animal reservoirs. No polioviruses have been isolated from nonprimates in nature, and the success of wild poliovirus eradication in the Americas indicates no previously unsuspected source of wild poliovirus in wild or domestic animals. Nonhuman primates were used extensively in early studies on polio infection and revealed that chimpanzees and some monkeys are susceptible to oral poliovirus infection, but to a lesser degree than humans (26). The low densities of such higher primate populations in the wild, their lower susceptibility to poliovirus infections, and the lack of evidence for a persistent carrier state suggest that these animals are unlikely to sustain continued transmission (27).

Environmental persistence. Fecal contamination is ubiquitous. Wild polioviruses and OPV viruses have been isolated from floors and furniture in households and daycare centers containing infected individuals (28, 29). The viruses can remain viable in contaminated soil, sewage, or water for some time, and even sewage treatment does not inactivate the full virus content (30). The half-life for survival of polioviruses, as measured by many studies on attenuated strains, depends on environmental conditions and is adversely affected by low relative humidity (31), low moisture content, high temperature, low pH, low organic content, and high bacterial growth (32). Differential environmental survival associated with seasonal changes in temperature and humidity may explain the seasonality associated with the prevalence of natural enterovirus infections (2, 33, 34).

Temperature is the most important determining factor for poliovirus survival, with 90 percent inactivation in a variety of surface waters seen only after 133–259 days at -20°C, 17.7–24.7 days at 1°C, and 3.8–6.7 days at 22°C, under laboratory conditions (32). Irrigation of crops with treated sewage effluent is practiced increasingly in industrialized countries. The high organic content favors poliovirus survival, and viable virus has been recovered from crops 23 days after treatment by sludge or effluent irrigation (35). Land surface application of sewage effluent creates the potential for poliovirus to overwinter in ice resulting from the process (32). Viruses can migrate to potable groundwaters, where survival is greater (29 days for 90 percent inactivation at 12°C) than in surface waters (30). The use of "nightsoil" for fertilization is practiced in many developing countries, and the survival of polioviruses in untreated sewage is likely to be longer than in the treated sewage measured in many studies. These reports suggest that poliovirus would be rapidly inactivated under tropical conditions, but temperate environments with high levels of environmental contamination (perhaps in the Andes or in the Himalayas) could favor soil- and waterborne poliovirus as a source of infection for some months. The probability of poliovirus survival for as long as a year appears to be remote except under unusual conditions, for example, involving subfreezing temperatures.

Laboratories. Many laboratories maintain living polioviruses of wild or OPV origin, either as part of ongoing research activities, or unknowingly in stored clinical or environmental samples. It will be a major task for the Global Poliomyelitis Eradication Initiative to ensure that such sources are contained, or preferably removed entirely, by destruction of all polioviruses except reserve stocks required for vaccine production. These must be kept under the tightest security conditions, and this will be a difficult task. There will also be a need to maintain large stocks of vaccine as a contingency after the eradication of poliovirus. Because of the high levels of control and containment required for OPV, stockpiling of IPV may be considered preferable under some circumstances, despite its being less efficient than OPV in outbreak control. There will be the associated issue of vaccinating laboratory workers involved in the production and maintenance of vaccine strains. It has been suggested that IPV may be used for this purpose (36), but this will need to be considered carefully, as wild prototype IPV viruses have already been isolated from clinical samples in the Netherlands and France (37). Alternatively, if OPV is used there may be a need to enforce postvaccination quarantine of staff until they are no longer potential sources of infection to contacts. The absolute necessity for this measure will depend on the potential for such viruses to spread and persist in susceptible human populations, the issue addressed in this report.

The practical difficulties associated with identification and destruction of laboratory virus stocks must be appreciated. Too few people today know that the last case of smallpox occurred in England in September 1978 as the result of a laboratory "accident." The potential for one or another poliovirus to be released inadvertently from a laboratory at some time in the future is substantial. This subject has been discussed at length, and a plan of action has been written for implementation in 1998 (36). This describes procedures for laboratory containment of wild polioviruses, which will be extended to OPV viruses after the cessation of OPV vaccination. In addition, it has been recommended that the World Health Organization seek agreement ultimately to destroy all poliovirus strains and potential infectious materials. Consideration may need to be given in the recommendations to materials that potentially harbor OPV viruses.

### Transmissibility of polloviruses—empirical evidence

The ability of OPV viruses to persist within semiimmune populations has been assessed by population/environmental surveillance following large-scale immunization of whole populations or targeted groups. Such studies form the basis of our current knowledge regarding the transmissibility of OPV viruses and are discussed briefly here. Few studies were carried out on the transmissibility of wild virus strains, and those available were considered for comparative purposes. It should be noted that, while these studies were carried out under both good and poor hygiene conditions, many are from communities in the United States and Russia and may not be wholly representative of the situation in developing countries.

Transmission to contacts. When OPV was introduced in the late 1950s, there were several investigations of the spread of OPV virus strains to direct and indirect contacts. Vaccinees were fed either monovalent, bivalent, or trivalent OPV and monitored for duration of fecal, and sometimes pharyngeal, excretion and virus titer (28, 38-54). In addition, unvaccinated siblings and other contacts were monitored for evidence of infection (fecal excretion or seroconversion) with the virus type fed. These studies varied in design, using Sabin, Cox, and/or Koprowski strains, employing different administration regimens and dosages, and recruiting individuals with different immune backgrounds. We have reviewed much of the published literature on such studies, using manual searching of a poliomyelitis abstracting journal (1951-1962), reference lists of publications, and computer-based searches of the MEDLINE reference database (1966-1998). The review may have missed a few non-English language publications but should not be otherwise biased. OPV now contains Sabin strains exclusively, so we excluded studies on Cox and Koprowski strains when estimating parameters. As IPV was available prior to the use of OPV, many investigations of the spread of OPV viruses were carried out in populations with prior IPV exposure. Though the effect of

IPV on subsequent excretion was a point of contention at the time many of these studies were carried out, some studies still did not stratify their data according to prior IPV exposure. The main findings of the studies on Sabin-type OPV virus transmission and the few studies on spread of wild poliovirus (2, 34, 55–62) are summarized in table 1 (complete tabulation is in the original report and is available on request from the authors). It is clear that many properties of wild and OPV viruses that affect transmission are similar.

*Reversion to neurovirulence.* To assess whether the live attenuated viruses might exhibit increasing neurovirulence upon replication in the human gut, several investigators have assessed the virulence of excreted OPV viruses. This was done with monkey neurovirulence tests or in vitro tissue culture markers correlated with neurovirulence, such as temperature and pH sensitivity. Wild viruses are more robust than are OPV viruses, surviving at higher temperatures and lower pH levels in culture (26). There have been reports of reversion of OPV viruses toward wild-type phenotypes and increased monkey neurovirulence of vaccine progeny (63–67). More recent studies at a molecular level have shown clearly that OPV viruses undergo extensive mutation, including reversion back toward wild-type genotypes (4).

An important consideration of this reversion is whether neurovirulence increases with passage through the human gut. Dane et al. (68) showed a progressive increase in monkey neurovirulence over time, after multiplication in the human gut, although Sabin (69) had found no evidence for this. Increased neurovirulence of OPV viruses excreted by a contact, compared with that excreted by the initial vaccinee, was reported from one study (63), while another found no evidence for this in contacts (70). A comparison of consecutive (second- and third-degree) contacts of vaccinees (siblings and contacts of siblings) found similar phenotypic reversion frequencies in viruses excreted by vaccinees and contacts (41). To account for dosage effects during contact infections, Dane et al. (68) purified OPV type 2 viruses excreted by a vaccinee and fed different doses (1-3 log tissue culture infective dose (TCID<sub>so</sub>)) to homotypic seronegative children. This study showed an increase in neurovirulence after a second passage but a similar degree of

TABLE 1. Summary of findings from WT\* and OPV\* virus transmission studies, under various conditions

Wild polioviruses	OPV viruses			
<ul> <li>Most susceptible contacts were infected</li> <li>Risk of transmission is associated with duration and intimacy of contact</li> </ul>	<ul> <li>Not much transmission under conditions of high preexisting immunity, considerable spread under conditions of low population immunity</li> <li>Young children are responsible for most transmission</li> <li>Greater effect of SES on differential transmission than with wild poliovirus</li> <li>Previously "immune" contacts can be reinfected, but replication is reduced (shorter, intermittent duration)</li> <li>Reinfection is dose dependent</li> </ul>			
<ul> <li>Young children are responsible for most transmission</li> </ul>				
Differential transmission associated with SES*				
<ul> <li>Previously "immune" contacts can be reinfected</li> </ul>				
Seasonal transmission	<ul> <li>Contacts often showed abortive excretion</li> <li>Contacts more susceptible to reinfection than vaccinees</li> </ul>			
<ul> <li>Equal transmission of all three serotypes over time</li> </ul>	<ul> <li>NPEV* interference effects (OPV dominant at high doses)</li> <li>Type 2 (generally) most transmissible</li> </ul>			
<ul> <li>Heterotypic antibodies do not protect against infection</li> </ul>	<ul> <li>Intertypic interference effects (excretion of individual types was less with TOPV* than with Mono)</li> <li>(Transient) heterotypic rise in antibodies</li> </ul>			
<ul> <li>Probability of fecal excretion unaffected by IPV* (but could only measure infection by</li> </ul>	<ul> <li>IPV not fully protective against reinfection (but duration of excretion reduced)</li> </ul>			
excretion, because of high antibody levels) <ul> <li>Pharyngeal excretion reduced by IPV</li> </ul>	<ul> <li>Pharyngeal excretion reduced by IPV at high antibody levels</li> </ul>			
<ul> <li>Some herd immunity provided by IPV</li> </ul>	<ul> <li>Probability of pharyngeal excretion after IPV, proportional to dose</li> </ul>			

\* WT, wild type; OPV, oral polio vaccine; SES, socioeconomic status; NPEV, non-polio enteroviruses; TOPV, trivalent oral polio vaccine (all three serotypes); IPV, formalin-inactivated polio vaccine.

neurovirulence after a third passage through the human gut (contacts of "secondary vaccinees"). Studies have suggested that the rate of mutation of polioviruses within the gut of one immunodeficient individual (15) is similar to that observed during circulation of wild poliovirus through the population over a 10-year period (71). This might enhance the probability of reversion to neurovirulence in immunodeficient OPV recipients.

*Reversion to wild-type transmissibility.* No evidence is available on whether OPV viruses "revert" toward the level of transmissibility of wild polioviruses during serial human-to-human transmission. This is a difficult issue to investigate, as the molecular and genetic basis of poliovirus transmissibility is not understood. A small number of such serial transmission studies were carried out in Russia during the late 1950s, designed to identify progressive trends in neurovirulence. The limited published data on fecal excretion titers show no evidence of consistent or significant change in transmissibility, as assessed from the viral titers excreted, over nine serial transfers of type 1, 2, or 3 OPV viruses (72).

Transmission within the community. Several studies have detected secondary spread of OPV viruses by serologic monitoring of unvaccinated individuals in areas known to be free of wild-type viruses. A seroprevalence study among preschool children with no record of prior vaccination in Houston, Texas, and Detroit, Michigan, showed convincing evidence of secondary spread of OPV viruses (42 percent type 2 seroprevalence in unvaccinated children) (73). Another study among Italian gypsy populations with very low vaccine coverage revealed greater than 90 percent seropositivity to type 2 poliovirus, which has been virtually eradicated from Europe, irrespective of vaccination status (74). Evidence for secondary spread of OPV viruses was also shown in a study in Oman, where infants given only IPV showed double the seroconversion rate to type 2 (82 percent) after potential exposure to targeted OPV campaigns, compared with those studied before the campaigns (41 percent) (75). These studies do not reveal the full potential of transmission of OPV viruses, as the extent of exposure to OPV viruses in these OPV-unvaccinated individuals is not known. In addition, high levels of natural or vaccine-attributable herd immunity in these populations may have limited the ability of OPV viruses to circulate.

Poliovirus persistence was monitored by population fecal surveys and sewage sampling following vaccination trials and campaigns when OPV was first made available. Studies in Czechoslovakia (76) and Hungary (77) showed that OPV viruses did not circulate at detectable levels after 5 months. Interpretation of these data is made difficult by the fact that both the Czechoslovakian and Hungarian populations vaccinated with OPV had received IPV during the previous 2 years. A trial in Mexico also showed little persistence of OPV viruses (78), but the preexisting high levels of natural immunity and the consequent lack of susceptible individuals must have limited the ability of OPV viruses to persist.

Many of these earlier studies did not distinguish between wild and attenuated polioviruses, making the data difficult to interpret. Fortunately, data on circulation of OPV viruses, in the absence of wild poliovirus, are available because of the unique poliovirus vaccination schedule in Cuba. All OPV is administered during two annual vaccination campaigns with no other routine poliovirus vaccination, providing an opportunity to study the persistence of OPV viruses in the environment and in the population during the rest of the year. Monthly cross-sectional stool surveys were carried out on children less than 2 years old and detected polioviruses only during and immediately after the vaccination campaigns (79, 80). A subsequent serologic survey, carried out on unvaccinated infants born 1–9 months after the last annual national immunization day, confirmed a lack of exposure to poliovirus in those born 3 months after the national immunization day (81). Because of the interfering effect of maternal antibodies in the children below 6 months, it is unclear whether OPV (especially type 2) continued to circulate for more than 2 months, and more intensive fecal surveillance will be necessary to answer this question.

Administration of trivalent oral polio vaccine (all three serotypes) to more than 90 percent of the Finnish population in 1984 to control a type 3 wild poliovirus epidemic (82) gave an opportunity to study the persistence of OPV viruses under conditions of intense and sensitive environmental surveillance. Countrywide sewage sampling detected OPV viruses for up to 6 months after the vaccination campaign (82, 83). However, while wild poliovirus type 3 was able to spread because of insufficient type-specific immunity (because of a poorly immunogenic type 3 IPV), it is likely that high levels of immunity (induced by IPV against types 1 and 2 and induced by the wild virus against type 3) limited the subsequent spread of the OPV viruses. Trivalent oral polio vaccine was also used to control an epidemic of type 3 wild poliovirus in the Netherlands in 1992-1993 (84), but sewage sampling was not carried out intensely enough to allow conclusions on the duration of OPV virus circulation in this population.

These results indicate that OPV viruses persisted within these populations for a very few months, but they provide no evidence that OPV viruses can persist

indefinitely. However, all of these study populations had high proportions of immune individuals among those not targeted for vaccination. It is clear that levels of immunity sufficient to stop wild poliovirus circulation in outbreak control campaigns must also block OPV virus circulation. We also note that national immunization days have been found to produce higher seroconversion rates than are achieved by routine vaccination (85). This is likely to be due in part to the intensive effort and better cold-chain facilities but also to increased exposure to OPV viruses. For unvaccinated individuals, both the probability of exposure to virus and the average dose encountered will increase with the intensity of the campaign effort. Thus, effective transmissions are likely to occur rapidly, reducing the proportion of susceptible individuals available for continued transmission. These implications of massive simultaneous use of OPV should be considered if and when OPV vaccination is stopped.

Also relevant are those communities where OPV is not accepted, where there is no endemic transmission of wild poliovirus, and where there is potential for exposure to OPV viruses. Such communities have little natural protection, the susceptible proportion will have increased since the local eradication of wild poliovirus, and any introduced OPV viruses would have an opportunity to invade the population. This is analogous to the situation that will arise some years after the cessation of vaccination and may provide us with important insights. A seroprevalence study of the conservative Amish communities in the United States revealed that 89 percent of unvaccinated children, born since eradication of wild poliovirus in the United States, had antibodies to type 2 poliovirus (Roland Sutter, Centers for Disease Control and Prevention, Atlanta, personal communication, 1997). A similar survey among unvaccinated communities in Canada detected that approximately 70 percent had antibodies to types 1 and/or 2 (Philippe Duclos, Laboratory Center for Disease Control, Ottawa, personal communication, 1998). It is important to note that, in many of the communities that reject vaccination in the Netherlands, Canada, and the United States, some members may accept vaccination, thus introducing OPV viruses directly into the community via multiple sources. However, it is clear that, once introduced, OPV viruses can spread in poorly vaccinated communities with good standards of hygiene, as seen in the Netherlands in 1992 (86).

The absence of VAPP "epidemics" (87) might suggest that, if OPV viruses do persist within unvaccinated communities, they rarely show increasing neurovirulence as they are transmitted between individuals. However, with the high levels of OPV coverage currently being attained in much of the world, it is unlikely that there are many susceptible communities large enough to enable continued spread of OPV viruses among susceptible individuals. Such a situation could have arisen in Albania, where suboptimal seroconversion due to poor cold-chain conditions led to a situation of low population immunity in the 18- to 35-year age group (88). Mass OPV campaigns targeted at children less than 5 years old were undertaken in 1996 and could have resulted in spread of OPV viruses to the remainder of the poorly vaccinated population. However, a simultaneous epidemic of wild type 1 occurred among the susceptible (mainly young adult) population (88), and this may have inhibited transmission of the OPV viruses. There is much anecdotal "evidence" about the absence of OPV virus spread in communities with low vaccine coverage, but the lack of seroprevalence data, the low case-to-infection ratio expected, and the absence of fecal sampling in these populations make it difficult to interpret such information.

More intensive surveillance for VAPP in countries with poor vaccination coverage could be helpful to supplement data on its frequency among populations with low levels of immunity. However, VAPP is difficult to define under conditions of endemic or recent endemic wild-type transmission, as discussed by Andrus et al. (87). VAPP is defined by exclusion of paralytic cases with wild-type virus isolation or any epidemiologic connection with wild-type cases or outbreaks. This is a necessity, under the impetus of eradication, to prevent any possible wild-type cases from being overlooked as VAPP cases. Nevertheless, this definition of VAPP, and the exclusion of paralytic cases with OPV virus isolation but no known OPV exposure history, could lead to VAPP cases being missed. These cases are important for the issue of OPV spread, and such "noncontact" VAPP cases need to be monitored closely to clarify their origin (87).

Such evidence reveals that OPV viruses, especially type 2, are transmitted frequently to susceptible individuals in direct contact with vaccinees and possibly also via infected nonvaccinees. While population and environmental sampling show limited persistence of OPV viruses after large, targeted campaigns, this has been influenced by high existing levels of immunity, high vaccine coverage, and concurrent infection of contacts, reducing the potential for continued transmission of OPV viruses. These studies do not show whether OPV viruses might persist in the very different circumstances that would prevail in a post-polio vaccination world. It is therefore important to estimate, as far as possible, the potential for OPV viruses to persist within largely susceptible populations. For this we turn to theoretical measures of transmissibility such as the secondary attack rate, basic reproduction number  $(R_{a})$ , and critical population size.

#### Transmissibility of polio viruses—theoretical approach

Several tools have been developed to measure the transmissibility and the persistence potential of infectious agents. The most useful of these are the ("secondary") attack rate, the (basic) reproduction number, and the concept of critical population size. The secondary attack rate is typically defined as the proportion of individuals exposed to a "primary" case who go on to develop disease because of that exposure. A very different measure of transmissibility is provided by the reproduction number of an infection. This concept describes transmission not as a risk, or probability, but as an average *number* of transmissions per case. As this will depend upon the prevalence of immunity in the population, the reference statistic, called the *basic* reproduction number  $(R_n)$ , is defined in theoretical terms as the average number of (successful) transmissions of infection (secondary "cases") that would occur if a single infected individual were introduced into a totally susceptible population. Finally, the longterm persistence of an infectious agent in a population is not just a function of its  $R_0$ , but it will also depend upon the proportion of susceptibles available for infection (determined by the nature of the immune response and the dynamics of the host population). This leads us to consider the concept of a critical population size, defined as the minimum *total* (human) population size required for maintenance, in perpetuity, of an infectious agent (89).

In order to assess the transmissibility of OPV viruses, we have emphasized comparisons between estimates for wild poliovirus and OPV viruses. This is because of the nature of the available data and the fact that the demonstrable ability of wild viruses to spread and persist provides a baseline against which to compare the estimated transmissibility of OPV viruses. Of course, OPV viruses must be less transmissible than wild polioviruses, or they would have driven out the wild viruses. The problem is to evaluate how much less transmissible they are.

Secondary attack rate. Estimates of the risk of transmission of polioviruses require close serologic and virologic surveillance of infected individuals and their contacts. While few studies were carried out on the risk of wild poliovirus infection among defined susceptible contact groups exposed to known "primary" cases, several investigators have provided estimates of transmission of OPV viruses among contacts of vaccine recipients. Although some of these measures are similar to the classical secondary attack rate,

in many studies the exposed contacts were not confirmed as susceptible at the time the source was vaccinated. If some contacts had some prior immunity, the secondary attack rate estimates will be too low. Another problem is the inability to distinguish between secondary and tertiary, etc., transmissions, as the classical secondary attack rate should include only secondary cases in the numerator. This is difficult to ensure in studies of polioviruses, as the serial intervals for secondary and tertiary transmissions overlap (58, 90), with infected individuals being able to excrete viruses within 24 hours of infection and to be infectious for several weeks. If the numerator includes individuals infected as the result of a third generation or extrafamilial transmission, the secondary attack rate estimates will be too high. Finally, numerical estimates for secondary attack rates will depend upon the context in which they are measured. The degree of intimacy varies between a household, institution, or community, and even between households, and the secondary attack rate estimate will vary accordingly.

Published infection risk (secondary attack rate-like) estimates for wild and OPV viruses on naive and IPVimmune backgrounds range from below 10 percent to over 90 percent (tables available in full report). The range reflects many study design, behavioral, and environmental factors. The accumulated data gave the impression that the transmissibility of OPV viruses is less than that of wild polioviruses, although the wide confidence intervals around the attack rate estimates, and the different social contexts used (households, closed institutions, schools, communities), make it difficult to interpret these results reliably. However, many of these studies reported a limitation in the transmission of OPV viruses, in particular Fox (39, p. 24), who concluded that "the most notable feature ... was the rapid decline in momentum of vaccine virus spread and its essential termination while a high [our italics] proportion of susceptibles were still available." We note below that this residual susceptible proportion is in fact a key (inverse) correlate of transmissibility.

Basic reproduction number (R). To measure the reproductive capacity of wild poliovirus and OPV viruses, we must consider several factors. 1) We are interested in the transmission of infection, not incidence of disease, and thus the statistic counts infected individuals. 2) As with the secondary attack rates, the basic reproduction number for any infection will obviously vary according to circumstances (crowding, hygiene behavior, and so on). Despite the problems inherent in its definition, the measure is used widely in theoretical epidemiologic work. This wide use was encouraged when Dietz pointed out that the  $R_0$  for an infection may be estimated as the reciprocal of the pro-

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portion susceptible to infection, for a population in which the infection is at some stable endemic state (91). From its definition, we appreciate that, if the  $R_0$  for an infection is below unity, then the infection cannot persist in a totally susceptible population. On the other hand, if the  $R_0$  exceeds unity, then some transmission of the infection is expected.

Though  $R_0$  provides one of the most elegant concepts in infectious disease epidemiology, it should not be overinterpreted. It reflects an average and carries an underlying assumption of homogeneity and random mixing, which does not reflect the real world. Given the global scope of the question before us and the tremendous variety of epidemiologic contexts in the real world, we will need to consider carefully the effect of this heterogeneity on estimates of transmissibility.

The basic reproduction number  $(R_{a})$  for OPV viruses cannot be measured directly from the proportion susceptible in a situation of "endemic stability," as no population exists in which the immune proportion has been determined solely by *natural* spread of OPV viruses. However, stability did exist for wild poliovirus in many populations prior to the introduction of routine vaccination in the 1950s. Many studies of age- and type-specific poliovirus antibody prevalence were carried out prior to the introduction of vaccination in communities around the world (57, 92–104). By estimating the values of  $R_0$  for wild polioviruses and comparing the parameters of wild and OPV viruses that influence transmissibility, it may be possible to provide a rough relative estimate of the  $R_{0}$ for OPV viruses and therefore to assess their potential for spread in susceptible populations.

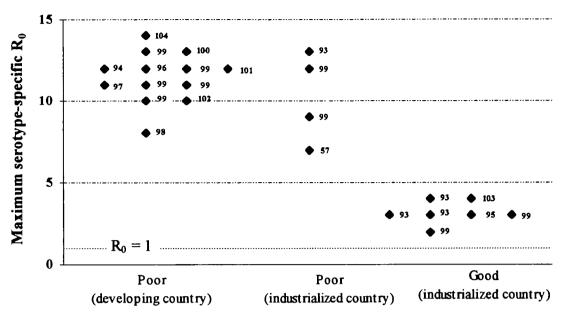
If an infection is endemic (and the population is demographically stable) then, on average, each infection leads to one subsequent infection. This implies that the "average" proportion immune =  $(R_0 - 1) / R_0$  =  $1 - (1 / R_{o})$ , and thus the susceptible proportion (S) = 1  $/R_{0}$  (105, 106). It may be noted that several authors have used the equation  $R_0 = L/A$  or  $R_0 = (1 + L/A)$  to derive estimates of  $R_0$ , where A is the average age at infection and L is the average life expectancy (reviewed by Dietz (107)). These formulas are shortcut approximations to estimate the proportion susceptible under assumptions of rectangular or exponential population structures, respectively (91, 105, 106), and they may not always be appropriate. Using published data on age-specific seroprevalence and available demographic data on the structure of the study populations (108, 109), we have estimated the proportions of each study population with and without type-specific antibodies against poliovirus and, hence, the  $R_0$  values for wild polioviruses under those conditions. The results show a reassuring consistency and are similar to those

derived by Eichner et al. (110, 111). We plot the maximum of the serotype-specific  $R_0$  values for each population in figure 1, excluding data for serotypes responsible for recent epidemics. One publication has derived estimates of the  $R_0$  of wild polioviruses higher than those presented here (for some of the same populations) (112), but these were based on the shortcut formula,  $R_0 = 1 + L/A$ .

Figure 1 shows that type-specific values of  $R_0$ ranged from 2 to 4 in areas of good hygiene and from 8 to 14 in areas of poor hygiene. This relation between transmission efficiency and standards of hygiene and sanitation is intuitively reasonable and has been noted by many authors (39, 57, 73, 99, 113). A high prevalence of diarrheal illness, reduced environmental survival in the dry season, and competitive interference by other enteroviruses may act to reduce the transmissibility of wild polioviruses in developing countries. It is important to note that the incidence of polio was not entirely "stable" in the populations in which these studies were carried out. Some had experienced recent epidemics, the studies were undertaken during different seasons, and the serotype distribution varied between years (59). Consequently, the differences in serotype-specific values of  $R_0$  may reflect in part the time since last introduction of particular serotypes rather than true differences in transmissibility. The estimates from areas of poor hygiene suggest that the three serotypes were transmitted to a roughly equal degree, and all three serotypes persisted within the same populations.

An important assumption of this method is that individuals with detectable antibodies (i.e., "humoral immunity") are immune to subsequent homotypic infection. However, many studies have shown that individuals with naturally induced immunity (defined by homotypic neutralizing serum antibodies at a dilution  $\geq 1:8$ ) can in fact be reinfected and excrete wild or attenuated poliovirus, especially after high challenge doses (2, 39, 59, 114, 115). If humoral immunity is not completely equivalent to gut immunity, it is important to assess to what extent an individual with humoral antibodies may still be "susceptible" to homologous reinfection and able to transmit the poliovirus.

A comparison of the total viral output from individuals with and without naturally and vaccine-acquired homotypic antibodies was carried out by challenging individuals with different prior exposures to poliovirus, with Sabin type 1 poliovirus (116). The study enables us to compare the *relative* potential of each group of individuals for further transmission. From table 2, we see that the proportion excreting, the duration of fecal excretion, and amount of virus excreted were all greater in the nonimmunes than in



#### General hygiene/sanitation conditions

**FIGURE 1.** Comparison of estimated  $R_0$  values for wild-type polioviruses from different studies (reference numbers given) in populations living under different sanitation and hygiene conditions. The highest serotype-specific  $R_0$  value has been used under the assumption that all serotypes have roughly the same potential for transmission in a given population, and any differences observed are due to epidemic patterns and the sequential transmission of serotypes through the population.

Immunologic status	Sample size	Proportion excreting Sabin 1	Mean excretion duration (days)	Mean log <sub>10</sub> virus titer (TCID <sub>50</sub> )/g of teces	Total viral output per person‡	Reduction In fecal virus excretion (%)
"Nonimmunes"	30	0.80	20.4	5.2	3.9 x 10 <sup>s</sup>	
Prior IPV*,§	31	0.74	12.3	4.1	1.7 x 10 <sup>7</sup>	96.00
Prior OPV§	33	0.37	4.6	2.2	4.1 x 10⁴	99.99
Prior wild type 1 infection§	19	0.37	5.0	2.2	4.4 x 10⁴	99.99
Immune to three types§	32	0.34	5.4	2.0	2.7 x 10⁴	99.99
Prior paralytic polio§	18	0.00				100

TABLE 2. Fecal virus excretion by groups with different previous exposure to wild or OPV\* viruses, after challenge with 6 logs of Sabin type 1†

\* OPV, oral polio vaccine; IPV, formalin-inactivated polio vaccine.

† Y. Z. Ghendon and I. I. Sanakoyeva. Acta Virol 1961;5:265-73.

‡ Assuming 150 g of feces per day.

§ All with high serum antibody titers against type 1 (1:64-1:256).

those with evidence of previous homotypic intestinal infection with wild or OPV viruses, or with prior IPV.

These data imply that a prior wild or OPV homotypic infection results in a reduction in excretion potential of approximately 99.99 percent. This reduction in virus output is considerable and provides support for the assumption that humoral antibody attributable to past *infection* is a reasonable surrogate for gut immunity with respect to reducing transmission. Consequently, we do not believe that our equation of humoral and intestinal immunity, implied in the method of deriving estimates of  $R_0$ , has led to serious error. If some seropositive individuals can be infected and significantly contribute to further transmission, the assumption will lead to an overestimation of  $R_0$ , that is, a conservative estimate for our purposes (discussed in original report).

These estimates of  $R_0$  for wild polioviruses are based on data collected several decades ago, and it is likely that hygienic conditions have improved in most industrialized countries since then. However, the documented spread of wild viruses in the Finnish outbreak of 1984 (82) and the outbreaks in unvaccinated groups in the Netherlands in 1978 and 1992 (86, 117), the United States in 1979 (118), and Canada in 1993 (119) show that the  $R_0$  of wild polioviruses is still greater than unity even in wealthier, industrialized countries.

Because of similarities in the epidemiology of hepatitis A and poliovirus prior to the introduction of poliomyelitis vaccination (103), current patterns of hepatitis A transmission (in the absence of vaccination) may provide an indication of the potential epidemiology of natural poliovirus infection under current conditions. Studies have revealed a marked reduction in the prevalence of hepatitis A antibodies in recent years, for example, in urban Hong Kong and Taiwan, where the socioeconomic status and conditions of sanitation have improved (120, 121). In contrast, studies in rural communities with poor hygiene have shown that extremely high exposure to hepatitis A still occurs (121, 122), indicating that, in these situations, hygiene and environmental sanitation conditions may be as favorable for the spread of poliovirus now as they were 40 years ago.

The greatest potential for persistence of OPV viruses is likely to occur in the poorer, "developing" country populations of the world. Our estimates show that  $R_{o}$ values of 10-15 prevailed in such countries for wild polioviruses, at the times these studies were carried out. We suspect that the hygienic conditions in many poor populations have not improved greatly in recent decades, in particular considering the growth of urban slums. It is possible that, in some populations of the world, perhaps urban shanty towns in Latin America, Africa, or Asia, the  $R_0$  for wild poliovirus is now greater than 15. However, in the absence of evidence to the contrary, we conclude, on the basis of this literature review and analyses, that the  $R_0$  of wild poliovirus is unlikely to exceed 20 for any populations of appreciable size today.

Several studies have shown that, under conditions of poor hygiene, seroconversion responses after OPV vaccination are generally lower in the tropics than seen in temperate countries (75, 123-126). This has been attributed to various factors, including high levels of maternal antibodies from boosted natural immunity in endemic situations (127), interference by concurrent enteroviral infections, diarrhea, poor nutritional status, and so on (123, 125, 126). Interference between OPV viruses and other enteroviral infections has been reported by many studies (28, 39, 40, 52, 53, 128-131), though others have reported no apparent interference in OPV "take" by preexisting enteroviral infections (42, 49, 124, 132). The higher seroconversion after IPV versus OPV vaccination in tropical situations (133) favors interference as an important influence. Our  $R_0$  estimates already take into consideration these various factors, including any interference that was occurring in the populations where these seroprevalence studies were carried out. Such interfering mechanisms are likely to affect transmission of wild as well as OPV viruses.

Extrapolation of  $R_0$  from wild to OPV viruses. To estimate how much lower the  $R_0$  is for OPV than for wild polioviruses, we examined in turn the several components of poliovirus transmissibility that are likely to affect  $R_0$ : infectious dose, duration of fecal (or pharyngeal) virus excretion, amount of virus passed in fecal excretion or pharyngeal secretions, and virus stability under various environmental conditions. The relative transmissibility of individual serotypes may not be identical for wild and OPV viruses. We have noted above that the transmissibility of types 1, 2, and 3 wild polioviruses appears to be similar, based on seroprevalence data, whereas studies of OPV viruses suggest that type 2 is more transmissible than the other serotypes.

It is not possible to determine the infectious dose of wild viruses for humans, but studies have been carried out on the minimum infectious dose of attenuated strains in humans. Early studies on Koprowski and Cox strains indicated that some, but not all, naive individuals could be infected with doses of just 1-2  $\text{TCID}_{so}$  (!) (134). The Sabin type 2 virus was shown to be infective in some individuals at doses of 2 log  $TCID_{cr}/g$  but was not infective to others at 3 logs. The latter finding may have been complicated by an effect of heterotypic priming at low doses (26). If we assume that the more neurovirulent, non-Sabin strains were less attenuated, these results suggest that larger infective doses are required for the more attenuated strains. There is evidence that hygiene conditions have a greater effect upon the dissemination of OPV viruses than on wild polioviruses (38, 39, 45) (although this may have been confounded by effects of IPV-induced immunity on pharyngeal excretion). If so, this may reflect at least in part that a higher dose of OPV virus than of wild poliovirus is required to initiate human infection.

Analyses of data on the duration of excretion (2, 48, 50, 53, 62, 135-137) indicate that this may be marginally less for OPV than for wild polioviruses (details in main report) (138).

The limited data available on fecal and pharyngeal viral titers indicate that these were roughly comparable for wild and OPV virus infections. This was the conclusion of a review by Melnick and Rennick (139), though Sabin (26) had reported that attenuated strains generally yielded 6  $\log_{10} \text{TCID}_{50}$  of virus per gram of feces, but that the current Sabin type 1 regularly yielded 100-fold lower peak titers. In addition, an association between neurovirulence and tissue culture

plaque size has been shown, with more attenuated strains producing smaller plaques (26, 65), suggestive of reduced multiplication by OPV viruses.

There are few comparative data on the environmental stability of wild and OPV viruses. Although temperature and pH sensitivity markers of neurovirulence suggest that wild polioviruses are more robust than the OPV viruses, the only comparative study reported that attenuated strains were more resistant to the effects of chlorination and adsorbed better to different soils than virulent strains (140). As reversion toward wild-type phenotypes on passage through the human gut may reduce any differences in survival capacity, there is a need for studies on the environmental survival of vaccine virus progeny.

Finally, we need to consider the effect of competition or interference by other enteroviruses that may obstruct transmission of OPV viruses. If OPV viruses are less able than wild-type viruses to resist such competition, this will introduce a differential infectiousness/transmissibility between OPV and wild-type viruses. Evidence of OPV competitive advantage against wild-type virus comes from studies where OPV was used to stop epidemics and was seen to replace the wild-type virus when wild-type titers were low, but not when wild-type titers were high (129, A review by Reichler and Patriarca (141) 131). describes situations in which outbreak control with OPV was not successful. All these failures were related to late initiation of vaccination (sometimes in association with low coverage), and they are therefore consistent with the already established virus being the more successful competitor. Competition studies in tissue culture have confirmed that the outcome of interference is dependent on the relative amounts of each virus type present (142). This suggests that competition can work both ways, depending on the growth phase of the established infection when a second infection is encountered, and provides no supporting evidence for OPV viruses being less competitive than wild poliovirus.

Interference among serotypes of OPV viruses has been observed (47, 53, 78, 143, 144), with type 2 generally producing higher seroconversion rates, despite being less immunogenic than type 1 (40, 47). This is clearest during the peak enterovirus transmission season, when seroconversion to types 1 and 3, but not type 2, tends to decline (123, 126). In vitro competition studies have reported a delay in multiplication of types 1 and 3 in the presence of type 2 (142). The relative amounts of types 1, 2, and 3 in trivalent oral polio vaccine have been adjusted to account for this, with ratios of 10:1:3 recommended (10:1:6 in developing countries (145)). There is evidence that the type 2 Sabin strain is more robust than the other serotypes, and this may be because it was a primary isolate, unlike types 1 and 3, which were products of laboratory selection from "wild" strains (26). On administration of trivalent OPV, type 2 is excreted most (48, 51), appears to be most transmissible to contacts (41, 47, 73, 74, 146), and is most prevalent in environmental surveys (83, 147). This heterogeneity in transmissibility of OPV serotypes will be an important consideration when assessing the probability of persistence of any single OPV strain.

In summary, these data indicate that OPV viruses are no fitter than wild-type strains of poliovirus in any single characteristic (except perhaps environmental stability?). This is of course to be expected, as wild poliovirus strains are ipso facto the result of selection under human transmission conditions. The crucial issue of the magnitude of the overall "fitness" difference between OPV and wild strains is very difficult to assess. It may be twofold, tenfold, or even greater. Though a precise relative estimate is not accessible, an important point for this discussion is that the difference in transmissibility may in fact be only twofold or fivefold. The only estimate we have noted for this differential is in a paper by Eichner and Dietz (111), who assume the  $R_0$  of OPV viruses to be 25 percent that of wild viruses (using estimates from 15 to 35 percent). However, even if the transmissibility of OPV viruses were 10-15 times less than that of wild polioviruses, it is likely that the  $R_0$  of some OPV viruses, in particular type 2, would be greater than unity in some circumstances. Thus, in theory, OPV viruses could persist indefinitely by person-to-person transmission if introduced into a totally susceptible population under poor hygiene conditions.

Accumulation of susceptibles. While the  $R_0$  measure provides an appropriate baseline measure of "absolute" transmissibility, it does not reflect actual (average) numbers of transmissions that occur in populations with some immunity. In theory, and assuming random mixing, the effective number of transmissions of an infection should be measured by  $R_0$  multiplied by the proportion susceptible in the population ( $R_0 \times S$ ). This actual number,  $R_n (= R_0 \times S)$ , is sometimes called the *effective* or net reproduction number (148).

In considering whether OPV viruses will persist in a community after cessation of vaccination, it is important to appreciate that the proportion susceptible in most of the world's populations is now very low. This is a function of high vaccine coverage in recent years (a *global* average of 82 percent for third doses of trivalent oral polio vaccine among infants in 1996 (149), in addition to widespread campaigns among children in many countries) and some "natural" immunity (attributable to wild poliovirus) among older individuals. It is therefore likely that the *average* number of successful transmissions  $(R_n)$  of OPV viruses per OPV-infected individual is currently appreciably less than unity in virtually all populations of the world. It is for this reason that transmission of OPV viruses appears to be time limited in all circumstances where it has been measured. This will be the situation if and when routine polio vaccination ceases.  $R_n$  will then begin to increase, as the proportion of susceptible individuals begins to rise immediately upon cessation of vaccination.

To evaluate the implications of this change, we have estimated the increase in the proportion susceptible in human populations. The method assumes that individuals are born with a prevalence of maternally derived antibodies equivalent to the prevalence of immunity in the adult population, and that this maternal immunity has a half-life of 1 month (136, 150) (it thus has little effect on the results). Two scenarios were modeled, that of lifelong mucosal immunity (figure 2) and that of mucosal immunity lost at a rate of 1 percent per annum. Using population-specific birth, death, and infant mortality rates, and starting with different susceptible proportions, one can estimate the time until the (potential) net reproductive number ( $R_0 \times S$ ) should reach unity, for a range of OPV serotype-specific  $R_0$ values. Once  $R_{1}$  exceeds unity, we expect any individual infected with OPV viruses to transmit the infection, on average, to at least one other individual. The figure thus illustrates the window of time, after cessation of vaccination, before conditions appropriate for endemic persistence of OPV viruses would be reached given a variety of assumptions.

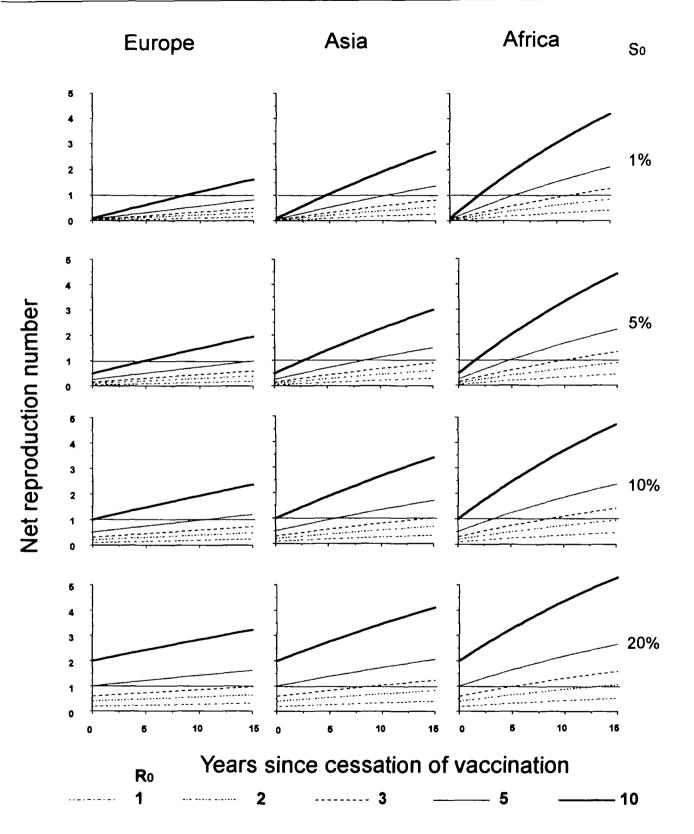
These analyses suggest that, for example in Africa, if the  $R_0$  for OPV viruses were 3 and immunity did not wane, it would take approximately 11, 10, 8, or 5 years for the potential  $R_n$  to exceed unity, if 1 percent, 5 percent, 10 percent, or 20 percent of the population were susceptible at the time vaccination ceased. These intervals would be shortened to 8.5, 8, 7.5, or 4 years, respectively, if immunity waned at 1 percent per annum (see original report for details). The "window" period would be appreciably shorter if the true  $R_0$  for OPV virus were as high as 5 or 10.

Such estimates should not be overinterpreted. They represent averages and imply isolated populations in which people mix at random. In particular, these analyses do not include age differences in contact patterns. *This may be important, as young children are generally found to have the highest secondary attack rate values* (2, 34, 43, 55, 57, 61), *and the increased susceptible proportion after cessation of vaccination will be precisely in these high-risk age groups.* Despite the several assumptions on which they are based, these results provide not unreasonable descriptions of possible scenarios.

An important consideration when predicting the rate of accumulation of susceptibles is the persistence of mucosal immunity. This is difficult to measure under current conditions of intensive vaccination, because of the potential for repeated infection by circulating OPV virus. A study by Nishio et al. (151) reported an increase in antibody levels over a period of years among Japanese vaccinees, suggestive of boosting by natural infection with OPV viruses. Although there are data on the persistence of humoral immunity in the absence of boosting (natural or vaccine related), there are no data on the persistence of mucosal immunity under these circumstances. Levels of mucosal immunity can be measured by challenge with OPV viruses (116, 144, 151, 152), and such studies have shown an inverse relation between levels of serum antibodies and viral excretion. This is clearly dependent on the mode of antibody induction, as antibodies attributable to prior intestinal infection by wild or OPV viruses are associated with much greater levels of intestinal immunity than are IPV-induced antibodies of the same level (see table 2).

Studies suggest that IPV-induced antibody levels are associated with reduced fecal excretion rates only at high antibody levels  $(\geq 1:128)$  (34, 62), while infection-induced antibodies are associated with reduced fecal excretion rates at lower levels  $(\geq 1:8)$  (151). There is an inverse association between antibody level and excretion rate (34, 54, 153, 154), even for maternally derived antibodies (136, 155). On the basis of such data, we can infer the persistence of intestinal immunity from information on the duration of detectable serum antibodies. The most appropriate data come from a serologic survey of Eskimo populations (92). There was evidence that polioviruses had not been maintained in this population, with a lack of antibodies to type 2 in individuals below 19 years old and a lack of antibodies to types 1 and 3 in individuals below 30 and 40 years old, respectively. Despite this lack of exposure for 20 years, serum antibody to type 2 persisted at a titer  $\geq 1:100$  in individuals aged 20-40 years old, after which antibody levels declined with age (92). Assuming that high levels of antibodies equate to gut protection, this suggests that intestinal immunity was maintained for at least 20 years in the absence of boosting, in younger adults, and this is supported by other such studies (99, 102, 156).

Critical population size. Even if an OPV strain were introduced into a population in which infection incidence increased (i.e., in which  $R_n = S \times R_0 > 1$ ), it does not necessarily follow that transmission would persist indefinitely. Persistence implies a sufficient



**FIGURE 2.** Estimates of the (potential) net reproduction number for oral polio vaccine (OPV) strains in relation to the time since cessation of vaccination in different populations, assuming different proportions susceptible  $(S_0)$  when vaccination is stopped. The model uses population-specific birth, mortality, and infant mortality rates (see text) and explores different estimates of the basic reproduction number  $(R_0)$  for OPV strains. Maternal immunity is assumed to last, on average, for 1 month, while naturally or vaccine-acquired immunity is lifelong.

supply of susceptibles and therefore depends on birth rates and total population size. The difficulty in estimating a critical population size for polio arises from the fact that it is a complex function of epidemiology, behavior, hygiene, and demography, each of which varies greatly among different "populations." We know from serologic studies that isolated populations of a few hundred individuals cannot maintain wild polioviruses permanently, as evidenced by the seronegativity of individuals born after periodic epidemics (70, 92, 157) due to introduced virus.

There are no seroprevalence data from large island populations, which might allow us to estimate how large an isolated population must be to support persistence of (wild) polioviruses, but it is likely to be on the order of several hundred thousand. Eichner et al. (110) used a simple simulation model to estimate the critical population size for wild poliovirus. They noted its dependence upon virus transmissibility  $(R_{o})$  and even more on population turnover (e.g., birth rate) and estimated that wild poliovirus could persist in unvaccinated populations of 500,000 under conditions of good hygiene ( $R_0 = 5$ ) and low birth rate (i.e., industrialized countries). With higher levels of population growth (i.e., developing countries) and conditions of poor hygiene ( $R_0 = 12$ ), a virus could persist in populations as low as 100,000. Their basic model assumed homogeneous mixing and no seasonality, and the inclusion of heterogeneous contact patterns improved persistence of the virus, thus reducing the critical population size.

Whether the critical population size for OPV viruses would be much greater than this is an even more complicated issue, as the efficiency of transmission is not a simple determinant of critical population size. Infections that spread very efficiently may exhaust susceptibles rapidly and may therefore need large population sizes or geographic heterogeneity for maintenance. Alternatively, poorly transmissible infections may only be able to persist in a large and dense population. We are thus unable to provide a convincing estimate of the theoretical minimum human population size necessary for persistent transmission of OPV viruses. On the other hand, while an interesting concept, the critical population size is of questionable relevance for the immediate practice of poliovirus eradication, for the simple fact that most human populations are heavily vaccinated (whereas the critical population size refers by definition to unvaccinated populations) and are not isolated. We consider the heterogeneity and interaction of contemporary human populations to be more important for the ultimate persistence of OPV viruses than the possibility that some isolated population will independently maintain these viruses for a long time.

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#### Predicting the future

We must do our best to anticipate what will/might happen if the current use of OPV succeeds in global eradication of wild polioviruses, and this is followed by cessation of polio vaccination. Precisely what happens will depend upon the strategy used for the discontinuation of vaccination, in particular, whether it is geographically phased out or simultaneous, whether it involves discontinuation of all OPV or sequential stopping (i.e., dropping one serotype from the vaccine first), and whether special end-stage vaccination strategies are used (e.g., global OPV or IPV national immunization days, or wide-scale use of IPV). We consider here the simplest scenario, that wild polioviruses are in fact eradicated (at least from human populations, if not laboratories) and that wide-scale use of OPV continues at essentially current levels until some date, for example, December 31, 2005, when it is stopped simultaneously throughout the world. All available evidence suggests that OPV viruses would continue to circulate and to be detectable in the environment of most populations (especially in sewage) for several months after cessation of vaccination. This duration will vary among populations but, given the variety of epidemiologic contexts, some viable virus may persist within certain closed communities and/or environments for at least a year after cessation of vaccination.

What happens next will depend on several factors: the initial proportion and distribution of susceptibles. the rate of increase of susceptibles (a function of birth rates and possibly loss of immunity), opportunities for transmission within the population (a function of hygiene, behavior, season, age distribution, social structure...), and opportunities for transmission among populations (a function of travel, proximity, migration, and population upheaval). Our analyses of  $R_{n}$  indicated that in any single population it is unlikely that transmission of OPV viruses would increase  $(R_{n})$ exceed unity) for at least 3 years after cessation of vaccination. These analyses assume a very high prevalence of immunity at the time of cessation of vaccination, and they do not include either the age heterogeneity within populations, the variance expected among populations, or the opportunities for transmission among them. Real world population heterogeneity provides a network of "high-risk" populations (generally associated with poor hygiene conditions and low vaccine coverage), such as the networks that enabled spread of wild poliovirus to Amish communities of the United States in the 1970s. Considering the immense heterogeneity of populations and transmission patterns, as well as the low detectability of infections with OPV viruses (a function of the extremely low case:infection ratio), there will be considerable opportunity for undetected transmission of OPV viruses, particularly in large, urban populations of Latin America, Asia, and Africa. On the basis of current knowledge, we cannot be certain that OPV would not continue to circulate somewhere or in some complex of populations for the 2–3 years prior to the virus being able to persist in a single population.

OPV virus may disappear from circulation within a year or two of cessation of vaccination. Another question arises when considering the situation several years after vaccination ceases, when the world will contain an appreciable proportion of poliovirus-susceptible individuals, as a consequence of births and possibly also a decline in immunity. As the prevalence of susceptible individuals increases toward unity, the effective (at least potential) net reproductive number approaches  $R_{0}$ . The available data suggest that the  $R_{0}$ for OPV viruses is likely to be greater than unity in some poorer populations, though perhaps not in the industrialized countries. There is therefore the possibility that, were OPV viruses to be successfully eradicated but then reintroduced into a poor, developing country many years later, they would persist there. This might be associated with very little paralytic ("VAPP") disease, perhaps with a morbidity incidence as low as one case per million. On the other hand, continued person-to-person transmission could lead to selection of the viruses toward wild-type properties associated with increased transmissibility and virulence. Such selection is likely to occur, though its rate and force cannot reliably be forecast.

Potential scenarios come to mind. An old vial of OPV is discovered in a refrigerator somewhere and is administered by someone not understanding the potential implications. Or an immunocompromised child in a developed country continues to excrete OPV viruses for several years after cessation of global vaccination. S/he then infects a friend, who travels to Rio de Janeiro (Bombay, Nairobi ...) while excreting poliovirus. Or, in the year 2010, a professor of parasitology in a school of public health examines stool specimens collected some years before and stored at -20°C. The laboratory does not use high containment for such procedures .... Such scenarios are not implausible. The probability of any one's occurring may be small, but the probability of something like them happening is not negligible. The plan of action for containment of polioviruses will need to encompass such eventualities (36).

#### Summary

The question of whether OPV viruses will (could, might ...) persist after cessation of vaccination does not admit a simple answer. Experience to date shows

that live OPV viruses will persist in most populations for at least several months after cessation of OPV vaccination, by a combination of environmental persistence and direct person-to-person transmission. Given the high levels of immunity that prevail in most populations because of extensive immunization coverage in recent years, the incidence and prevalence of OPV infections will decline rapidly and would probably not persist in any "single" population. However, given the variety, heterogeneity, interconnectedness, and sheer number of human populations, the possibility cannot be excluded that OPV viruses could succeed in persisting for several years, somewhere, in one or another population network. This may be associated with very little characteristic disease and may be exceedingly difficult to detect. The speculative nature of such predictions should be emphasized. The attempt to predict highlights important gaps in our knowledge.

A different situation will arise several years after cessation of vaccination, as the susceptible proportion increases through births, possibly enhanced by the waning of immunity in older individuals no longer exposed to poliovirus. Available information indicates that OPV viruses, in particular the Sabin type 2, would be able to persist indefinitely if introduced into a totally susceptible population living under conditions of poor hygiene. Transmission has generally been found to be most efficient among young children, and the accumulation of large numbers of susceptibles in this age group, soon after cessation of vaccination, will favor conditions for continued transmission. Thus, a crucial question is whether OPV viruses can persist "by luck" for a few years before conditions arise that would effectively ensure their persistence. The probability of this occurring is extremely difficult to assess a priori, but logic may point toward the wisdom of introducing a "2-less" OPV vaccine and monitoring closely what happens to the type 2 strain, before cessation of vaccination altogether (158, 159).

Another factor that is difficult to predict is the likelihood that OPV strains would revert to wild-type transmissibility, once "left" to themselves in a world with an increasing density of susceptibles and in the absence of competition from large numbers of recently administered vaccine viruses. Given that the wild- and vaccine-type viruses differ by only a few point mutations, such reversion is possible, but our ignorance of the molecular and genetic determinants of transmissibility does not allow us to quantify a priori the likelihood or extent to which it may occur.

Even if continued transmission of OPV strains is broken entirely, a danger exists for reintroduction of one or another poliovirus, most probably OPV, mediated through long-term excretion of viruses by some immunocompromised individual, or through accidental contamination or infection from stored vaccine or clinical or fecal material. The likelihood of such introductions will decrease with time, but the potential consequences increase with time, because of increasing numbers and proportions of susceptible individuals and therefore increased probability that any introduced virus would persist.

We conclude that there is a risk that OPV viruses will persist and that such persistence could occur in a variety of ways. Important questions remain unanswered, and research is needed to assess the implications for OPV virus persistence. Issues requiring the most immediate attention are those of long-term excretion by immunodeficient individuals, especially those with HIV and AIDS; the ability for OPV viruses to spread and persist in communities with low seroprevalence; the risk of reversion to wild-type transmissibility; environmental survival and potential reservoirs of OPV virus; duration of mucosal immunity; and the prevalence of viable poliovirus in stored samples. Careful consideration must be given to the implications of various options for phasing out vaccination and to implementing a research agenda that will provide appropriate information to guide policy, to ensure a successful end to this important public health endeavor.

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#### References

- 1. World Health Organization. Global eradication of poliomyelitis by the year 2000. Resolution 41.28, agenda item 12, pp 1-3, of the 41st World Health Assembly, Geneva, Switzerland, May 13, 1988.
- 2. Gelfand HM, LeBlanc DR, Fox JP, et al. Studies on the development of natural immunity to poliomyelitis in

Louisiana. II. Description and analysis of episodes of infection observed in study group households. Am J Hyg 1957; 65:367-85.

- Minor PD. The molecular biology of poliovaccines. J Gen Virol 1992;73:3065–77.
- Dunn G, Begg NT, Cammack N, et al. Virus excretion and mutation by infants following primary vaccination with live oral poliovaccine from two sources. J Med Virol 1990;32: 92-5.
- Abraham R, Minor P, Dunn G, et al. Shedding of virulent poliovirus revertants during immunization with oral poliovirus vaccine after prior immunization with inactivated polio vaccine. J Infect Dis 1993;168:1105-9.
- Fiore L, Pierangeli A, Lombardi F, et al. Antigenic and biochemical characterization of poliovirus type 2 isolated from two cases of paralytic disease. Intervirology 1987;27:196– 204.
- Lipskaya GY, Muzychenko AR, Kutitova OK, et al. Frequent isolation of intertypic poliovirus recombinants with serotype 2 specificity from vaccine-associated polio cases. J Med Virol 1991;35:290-6.
- Furione M, Guillot S, Otelea D, et al. Polioviruses with natural recombinant genomes isolated from vaccine-associated paralytic poliomyelitis. Virology 1993;196:199–208.
   World Health Organization. Primary immunodeficiency dis-
- World Health Organization. Primary immunodeficiency diseases: report of a WHO scientific group. Clin Exp Immunol 1997;109(suppl 1):1-28.
- 10. Wright PF, Hatch MH, Kasselberg AG, et al. Vaccine-associated poliomyelitis in a child with sex-linked agammaglobulinaemia. J Pediatr 1977;91:408–12.
- 11. Sutter RW, Prevots DR. Vaccine-associated paralytic poliomyelitis among immunodeficient persons. Infect Med 1994;11:426–38.
- MacCallum FO. Hypogammaglobulinaemia in the United Kingdom. VII. The role of humoral antibodies in protection against and recovery from bacterial and virus infections in hypogammaglobulinaemia. Spec Rep Ser Med Res Counc (G B) 1971;310:72–85.
- Hara M, Saito Y, Komatsu T, et al. Antigenic analysis of polioviruses isolated from a child with agammaglobulinemia and paralytic poliomyelitis after Sabin vaccine administration. Microbiol Immunol 1981;25:905–13.
- Savilahti E, Klemola T, Carlsson B, et al. Inadequacy of mucosal IgM antibodies in selective IgA deficiency: excretion of attenuated polio viruses is prolonged. J Clin Immunol 1988;8:89–94.
- Prolonged poliovirus excretion in an immunodeficient person with vaccine-associated paralytic poliomyelitis. MMWR Morb Mortal Wkly Rep 1997;46:641-3.
- Baumgart KW, Britton WJ, Kemp A, et al. The spectrum of primary immunodeficiency disorders in Australia. J Allergy Clin Immunol 1997;100:415-23.
- Fasth A. Primary immunodeficiency disorders in Sweden: cases among children, 1974–1979. J Clin Immunol 1982;2: 86–92.
- Thorsteinsson L, Ogmundsdottir HM, Sigfusson A, et al. The first Icelandic family with X-linked agammaglobulinaemia: studies of genetic markers and immune function. Scand J Immunol 1990;32:273–80.
- Grumach AS, Duarte AJ, Bellinati-Pires R, et al. Brazilian report on primary immunodeficiencies in children: 166 cases studied over a follow-up time of 15 years. J Clin Immunol 1997;17:340-5.
- Ion Nedelcu N, Dobrescu A, Strebel PM, et al. Vaccine-associated paralytic poliomyelitis and HIV infection. (Letter). Lancet 1994;343:51-2.
- Ryder RW, Oxtoby MJ, Mvula M, et al. Safety and immunogenicity of *Bacille Calmette-Guerin*, diphtheria-tetanus-pertussis, and oral polio vaccines in newborn children in Zaire infected with human immunodeficiency virus type 1. J Pediatr 1993;122:697-702.
- 22. Kitayaporn D, Tansuphaswadikul S, Lohsomboon P, et al. Survival of AIDS patients in the emerging epidemic in

Bangkok, Thailand. J Acquir Immune Defic Syndr Hum Retrovirol 1996;11:77-82.

- 23. Morgan D, Maude GH, Malamba SS, et al. HIV-1 disease progression and AIDS-defining disorders in rural Uganda. Lancet 1997;350:245-50.
- 24. Dunne MT, Ruskin HJ, Mulcahy FM. Survival with AIDS in Ireland. AIDS 1997;11:1281–90.
- 25. Centers for Disease Control and Prevention. Meeting on "Prolonged excretion of poliovirus in immune deficient persons." Atlanta: CDC, 1997.
- 26. Sabin AB. Present status of attenuated live-virus poliomyelitis vaccine. JAMA 1956;162:1589–96.
- Dowdle WR, Birmingham ME. The biologic principles of poliovirus eradication. J Infect Dis 1997;175(suppl 1): S286-92.
- Gelfand HM, Potash L, LeBlanc DR, et al. Intrafamilial and interfamilial spread of living vaccine strains of polioviruses. JAMA 1959;170:85/2039-94/2048.
- 29. Keswick BH, Pickering LK, DuPont HL, et al. Survival and detection of rotaviruses on environmental surfaces in day care centers. Appl Environ Microbiol 1983;46:813–16.
- Yates MV, Gerba CP, Kelley LM. Virus persistence in groundwater. Appl Environ Microbiol 1985;49:778-81.
- Hemmes JH, Winkler KC, Kool SM. Virus survival as a seasonal factor in influenza and poliomyelitis. Antonie Van Leeuwenhoek 1962;28:221-3.
- 32. Hurst CJ, Benton WH, McClellan KA. Thermal and water source effects upon the stability of enteroviruses in surface freshwaters. Can J Microbiol 1989;35:474–80.
- Yorke JA, Nathanson N, Pianigiani G, et al. Seasonality and the requirements for perpetuation and eradication of viruses in populations. Am J Epidemiol 1979;109:103–23.
- Fox JP, Hall CE. Poliomyelitis in Louisiana. In: Viruses in families: surveillance of families as a key to epidemiology of virus infections. Littleton, MA: PSG Publishing Company, Inc, 1980:93-151.
- 35. Tierney JT, Sullivan R, Larkin EP. Persistence of poliovirus 1 in soil and on vegetables grown in soil previously flooded with inoculated sewage sludge or effluent. Appl Environ Microbiol 1977;33:109–13.
- World Health Organization. Proposed global action plan and timetable for safe handling and minimum laboratory containment of wild polioviruses and potentially infectious materials. Geneva: World Health Organization, 1998. (WHO/EPI/ Gen/98.05).
- Mulders MN, Reimerink JH, Koopmans MP, et al. Genetic analysis of wild-type poliovirus importation into the Netherlands (1979–1995). J Infect Dis 1997;176:617–24.
- Fox JP, Hall CE. Experimental studies with vaccine strains of polioviruses. In: Viruses in families: surveillance of families as a key to epidemiology of virus infections. Littleton, MA: PSG Publishing Company, Inc, 1980:152–95.
- 39. Fox JP. The spread of vaccine strains of poliovirus in the household and in the community in southern Louisiana. (Abstract). In: Poliomyelitis: papers and discussions presented at the 5th International Poliomyelitis Conference, Copenhagen, Denmark, July 26–28, 1960. Philadelphia: JB Lippincott Co, 1960:24–5.
- 40. Benyesh-Melnick M, Melnick JL, Ramos Alvarez M. Poliomyelitis infection rate among Mexican children fed attenuated poliovirus vaccines. In: Proceedings of the 1st International Conference on Live Poliovirus Vaccines. Washington, DC: Pan American Sanitary Bureau, 1959: 272-85.
- 41. Benyesh-Melnick M, Melnick JL, Rawlis WE, et al. Studies of the immunogenicity, communicability, and genetic stability of oral poliovaccine administered during the winter. Am J Epidemiol 1967;86:112–36.
- 42. Sabin AB, Michaels RH, Spigland I, et al. Community-wide use of oral poliovirus vaccine: effectiveness of the Cincinnati Program. Am J Dis Children 1961;101:546–67.
- 43. Koprowski H, Plotkin S, Pagano J, et al. Behaviour of atten-

uated strains of poliomyelitis virus in relation to age, familial spread, and duration of immunity. In: Proceedings of the 1st International Conference on Live Poliovirus Vaccines. Washington, DC: Pan American Sanitary Bureau, 1959: 159-71.

- 44. Horstmann DM, Niederman JC, Riordan JT, et al. The trial use of Sabin's attenuated type 1 poliovirus vaccine in a village in southern Arizona. Am J Hyg 1959;70:169-84.
- 45. Fox JP. Factors influencing dissemination of wild and vaccine strains of poliovirus in Louisiana. In: Choumakov MP, ed. Oral live poliovirus vaccine. (Papers presented at the IVth Scientific Conference of the Institute of Poliomyelitis and Virus Encephalitis and the International Symposium on Live Poliovirus Vaccine, May 17-20, 1960). Moscow: Academy of Medical Sciences of the USSR, 1961:531-45.
- 46. Horstmann DM, Niederman JC, Paul JR. Attenuated type 1 poliovirus vaccine. Its capacity to spread from "vaccinees" within an institutional population. JAMA 1959;170:1–8.
- 47. Smorodintsev AA, Ilyenko VI, Kurnosova MM, et al. Virological and immunological characteristics of vaccinal infection in children inoculated per os with a live poliomyelitis vaccine made from the Sabin strains. In: Proceedings of the 1st International Conference on Live Poliovirus Vaccines. Washington, DC: Pan American Sanitary Bureau, 1959:312-23.
- Horstmann DM, Paul JR, Godenne-McCrea M, et al. Immunization of preschool children with oral poliovirus vaccine (Sabin). JAMA 1961;178:693-701.
- 49. Koroleva GA, Zhevandrova VI, Voroshilova MK, et al. Virologic and serologic survey of children in contact with those immunized with live poliovirus vaccine. In: Choumakov MP, ed. Oral live poliovirus vaccine. (Papers presented at the IVth Scientific Conference of the Institute of Poliomyelitis and Virus Encephalitis and the International Symposium on Live Poliovirus Vaccine, May 17-20, 1960). Moscow: Academy of Medical Sciences of the USSR, 1961: 584-7.
- 50. Drosdov SG, Shirman GA, Ashmarina EE, et al. Dynamics of excretion of the three types of vaccine virus in vaccinated children and their contacts. Investigation of properties of virus isolates. In: Choumakov MP, ed. Oral live poliovirus vaccine. (Papers presented at the IVth Scientific Conference of the Institute of Poliomyelitis and Virus Encephalitis and the International Symposium on Live Poliovirus Vaccine, May 17–20, 1960). Moscow: Academy of Medical Sciences of the USSR, 1961:165–73.
- 51. Zhevandrova VI, Koroleva GA, Voroshilova MK. The dynamics of vaccine strain excretion in oral immunization with trivalent live poliomyelitis vaccine. In: Choumakov MP, ed. Oral live poliovirus vaccine. (Papers presented at the IVth Scientific Conference of the Institute of Poliomyelitis and Virus Encephalitis and the International Symposium on Live Poliovirus Vaccine, May 17–20, 1960). Moscow: Academy of Medical Sciences of the USSR, 1961:174–80.
- 52. Ginter VX, Indulen MK, Kanel IA. Spread of vaccine virus in boarding children's institutions and households during immunization with live poliovirus vaccine. In: Choumakov MP, ed. Oral live poliovirus vaccine. (Papers presented at the IVth Scientific Conference of the Institute of Poliomyelitis and Virus Encephalitis and the International Symposium on Live Poliovirus Vaccine, May 17–20, 1960). Moscow: Academy of Medical Sciences of the USSR, 1961:588–90.
- Japan Live Poliovaccine Research Commission. Evaluation of Sabin live poliovirus vaccine in Japan. II. Clinical, virologic and immunologic effects of vaccine in children. Jpn J Med Sci Biol 1966;19:277–91.
- Glezen WP, Lamb GA, Belden EA, et al. Quantitative relationship of preexisting homotypic antibodies to the excretion of attenuated poliovirus type 1. Am J Epidemiol 1966;83:224-37.
- 55. Bodian D, Paffenbarger RS. Poliomyelitis infection in households: frequency of viremia and specific antibody response.

Am J Epidemiol 1954;60:83-93.

- Paffenbarger RS, Wilson VO, Bodian D, et al. The spread of poliomyelitis: an analysis of contact during epidemic periods. Am J Hyg 1954;60:63-82.
- 57. Fox JP, Gelfand HM, LeBlanc DR, et al. Studies on the development of natural immunity to poliomyelitis in Louisiana. I. Over-all plan, methods and observations as to patterns of seroimmunity in the study group. Am J Hyg 1957; 65:344-66.
- Isacson P, Melnick JL, Walton M. Environmental studies of endemic enteric virus infections. II. Poliovirus infections in household units. Am J Hyg 1957;65:29–42.
- 59. Gelfand HM, LeBlanc DR, Potash L, et al. Studies on the development of natural immunity to poliomyelitis in Louisiana. IV. Natural infections with polioviruses following immunization with a formalin-inactivated vaccine. Am J Hyg 1959;70:312-27.
- 60. Wehrel PF, Carbonaro O, Day PA, et al. Transmission of polioviruses. III. Prevalence of polioviruses in pharyngeal secretions of infected household contacts of patients with clinical disease. Pediatrics 1961;27:762-4.
- 61. Chin TDY, Marine WM, Hall EC, et al. Poliomyelitis in Des Moines, Iowa, 1959. The influence of Salk vaccination on the epidemic pattern and the spread of the virus in the community. Am J Hyg 1961;74:67–94.
- Marine WM, Chin TDY, Gravelle CR. Limitation of fecal and pharyngeal poliovirus excretion in Salk-vaccinated children. Am J Hyg 1962;76:173–95.
- 63. Melnick JL, Benyesh-Melnick M. Problems associated with live poliovirus vaccine and its progeny after multiplication in man. In: Proceedings of the 2nd International Conference on Live Poliovirus Vaccines. Washington, DC: Pan American Sanitary Bureau, 1960:12–27.
- 64. Voroshilova MK, Lavrova LK, Balayan MS, et al. Investigation of type III poliovirus strains, recovered from live virus vaccinees, in monkey tests and by titrations at 36°C and 40°C. In: Choumakov MP, ed. Oral live poliovirus vaccine. (Papers presented at the IVth Scientific Conference of the Institute of Poliomyelitis and Virus Encephalitis and the International Symposium on Live Poliovirus Vaccine, May 17-20, 1960). Moscow: Academy of Medical Sciences of the USSR, 1961:227-30.
- 65. Ghendon YZ, Khesin YE, Marchenko AT. Study of stability of the genetic characters of Sabin attenuated poliomyelitis virus strains. In: Choumakov MP, ed. Oral live poliovirus vaccine. (Papers presented at the IVth Scientific Conference of the Institute of Poliomyelitis and Virus Encephalitis and the International Symposium on Live Poliovirus Vaccine, May 17-20, 1960). Moscow: Academy of Medical Sciences of the USSR, 1961:456-76.
- 66. Robinson IA, Sheftel MA, Yurovetskaya AL, et al. Changes in the central nervous system of monkeys inoculated with type III strains isolated from feces of children vaccinated with oral live vaccine. In: Choumakov MP, ed. Oral live poliovirus vaccine. (Papers presented at the IVth Scientific Conference of the Institute of Poliomyelitis and Virus Encephalitis and the International Symposium on Live Poliovirus Vaccine, May 17–20, 1960). Moscow: Academy of Medical Sciences of the USSR, 1961:231–7.
- 67. Dane DS, Dick GWA, Briggs M, et al. Vaccination against poliomyelitis with live virus vaccines. 8. Changes in Sabin type I oral vaccine virus after multiplication in the intestinal tract. Br Med J 1961;2:269-71.
- Dane DS, Dick GWA, Briggs M, et al. Vaccination against poliomyelitis with live virus vaccines.
   Changes in Sabin type II oral vaccine virus after human passage. Br Med J 1961;2:259-65.
- 69. Sabin AB. Recent studies and field tests with a live attenuated poliovirus vaccine. In: Proceedings of the 1st International Conference on Live Poliovirus Vaccines. Washington, DC: Pan American Sanitary Bureau, 1959:14-33.

- Oker-Blom N, Strandstrom H, Eriksson AW. A small-scale trial with live poliovirus vaccine in an isolated island community. In: Proceedings of the 1st International Conference on Live Poliovirus Vaccines. Washington, DC: Pan American Sanitary Bureau, 1959:580-7.
- Kew OM, Mulders MN, Lipskaya GY, et al. Molecular epidemiology of polioviruses. Semin Virol 1995;6:401-14.
- 72. Klyuchareva TE. Changes in the neurotropic properties of attenuated strains of the poliomyelitis virus after lengthy circulation through the susceptible organism of children. In: Smorodintsev AA, ed. Living vaccine against poliomyelitis: a collection of articles from the Virology Department. (In Russian). Leningrad: Works of the Institute of Experimental Medicine, AMS, USSR, 1960.
- Chen RT, Hausinger S, Dajani AS, et al. Seroprevalence of antibody against poliovirus in inner-city preschool children. Implications for vaccination policy in the United States (see comments). JAMA 1996;275:1639-45.
- Aylward RB, Porta D, Fiore L, et al. Unimmunized gypsy populations and implications for the eradication of poliomyelitis in Europe. J Infect Dis 1997;175(suppl 1): S86-8.
- 75. World Health Organization. Combined immunization of infants with oral and inactivated poliovirus vaccines: results of a randomized trial in The Gambia, Oman, and Thailand. J Infect Dis 1997;175(suppl 1):S215-27.
- Zacek K, Adam E, Adamova V, et al. Mass oral (Sabin) poliomyelitis vaccination: virological and serological surveillance in Czechoslovakia, 1958–9 and 1960. Br Med J 1962;1:1091–8.
- Domok I, Molnar E, Jancso A, et al. Enterovirus survey in children after mass vaccination with live attenuated polioviruses. Br Med J 1962;1:743-6.
- polioviruses. Br Med J 1962;1:743-6.
  78. Sabin AB, Ramos-Alvarez M, Alvarez-Amezquita J, et al. Live, orally given poliovirus vaccine: effects of rapid mass immunization on population under conditions of massive enteric infection with other viruses. JAMA 1960;173: 1521-6.
- Mas Lago P, Louzara C, Beltran J, et al. Circulacion de poliovirus en la poblacion infantil de Cuba. (In Spanish). Bol Oficina Sanit Panam 1979;87:377-88.
- Ochoa EG, Mas Lago P. Epidemiological surveillance and control of poliomyelitis in the Republic of Cuba. J Hyg Epidemiol Microbiol Immunol 1987;31:381-9.
- Mas Lago P, Ramon Bravo J, Andrus JK, et al. Lessons from Cuba: mass campaign administration of trivalent oral poliovirus vaccine and seroprevalence of poliovirus neutralizing antibodies. Bull World Health Organ 1994;72:221-5.
- Hovi T, Huovilainen A, Kuronen T, et al. Outbreak of paralytic poliomyelitis in Finland: widespread circulation of antigenically altered poliovirus type 3 in a vaccinated population. Lancet 1986;1:1427-32.
- 83. Poyry T, Stenvik M, Hovi T. Viruses in sewage waters during and after a poliomyelitis outbreak and subsequent nationwide oral poliovirus vaccination campaign in Finland. Appl Environ Microbiol 1988;54:371-4.
- 84. van der Avoort HG, Reimerink JH, Ras A, et al. Isolation of epidemic poliovirus from sewage during the 1992-3 type 3 outbreak in the Netherlands. Epidemiol Infect 1995;114: 481-91.
- Richardson G, Linkins RW, Eames MA, et al. Immunogenicity of oral poliovirus vaccine administered in mass campaigns versus routine immunization programmes. Bull World Health Organ 1995;73:769-77.
- Conyn van Spaendonck MA, Oostvogel PM, van Loon AM, et al. Circulation of poliovirus during the poliomyelitis outbreak in the Netherlands in 1992–1993. Am J Epidemiol 1996;143:929–35.
- Andrus J, Aylward B, Hlady G. Vaccine-associated paralytic poliomyelitis (VAPP)—VAPP issues in endemic countries. (Technical consultation on the global eradication of poliomyelitis). Geneva, Switzerland: World Health

Organization, 1997. (EPI/POLIO/TECH.97/WP.10).

- Prevots DR, Ciofi degli Atti ML, Sallabanda A, et al. Outbreak of paralytic poliomyelitis in Albania, 1996: high attack rate among adults and apparent interruption of transmission following nationwide mass vaccination. Clin Infect Dis 1998;26:419-25.
- Black FL. Measles endemicity in insular populations: critical community size and its evolutionary implication. J Theor Biol 1966;11:207-11.
- Aycock WL, Kessel JF. The infectious period of poliomyelitis and virus detection. Am J Med Sci 1943;205: 454-65.
- 91. Dietz K. Transmission and control of arbovirus diseases. In: Ludwig D, Cooke KL, eds. Epidemiology. Philadelphia: Society of the Institute of Applied Mathematics, 1975: 104-21.
- Paul JR, Riordan JT, Melnick JL. Antibodies to three different antigenic types of poliomyelitis virus in sera from North Alaskan Eskimos. Am J Hyg 1951;54:275–85.
- Paul JR, Melnick JL, Riordan JT. Comparative neutralizing antibody patterns to Lansing (type 2) poliomyelitis virus in different populations. Am J Hyg 1952;56:232-51.
- Paul JR, Melnick JL, Barnett VH, et al. A survey of neutralizing antibodies to poliomyelitis virus in Cairo, Egypt. Am J Hyg 1952;55:402-13.
- 95. Melnick JL, Ledinko N. Development of neutralizing antibodies against the three types of poliomyelitis virus during an epidemic period. The ratio of inapparent infection to clinical poliomyelitis. Am J Hyg 1953;58:207-22.
- Paul JR, Horstmann DM. A survey of poliomyelitis virus antibodies in French Morocco. Am J Trop Med Hyg 1955;4: 512-24.
- Gelfand HM, Miller MJ. Poliomyelitis in Liberia. Prevalence of the disease, sero-immunity resulting from sub-clinical infection, and indications for prophylactic vaccination. Am J Trop Med Hyg 1956;5:791-6.
- Coriell LL, Schaeffer K, Felton HM, et al. A serologic and clinical survey of poliomyelitis in Caracas, Venezuela, and Galveston, Texas, and the response to Salk vaccine. Am J Public Health 1956;46:1431-8.
- Melnick JL. Studies on the serological epidemiology of poliomyelitis as an index in certain Caribbean islands, British Guiana, and Ecuador. West Indian Med J 1959;8:275-98.
- Fendall NRE. Poliomyelitis in Kenya—the 1960 epidemic and oral vaccine campaign. J Trop Med Hyg 1962;65: 245-55.
- Wai-Kwan C, Hay S. Poliomyelitis fecal and serological surveys in the Chinese population in Hong Kong in 1960. Am J Trop Med Hyg 1962;11:122-5.
- Olness KN, Halstead SB, Snitbhan R. Poliomyelitis in Laos, 1962–1963. Epidemic and immunity survey. J Pediatr 1966; 69:316–23.
- Gust ID, Lewis FA, Lehmann NI. Prevalence of antibody to hepatitis A and polioviruses in an unimmunized urban population. Am J Epidemiol 1978;107:54-6.
- Schonberger LB, Thaung U, Khi DK, et al. The epidemiology of poliomyelitis in Burma. Dev Biol Stand 1981;47: 283-92.
- 105. Anderson RM, May RM. Infectious diseases of humans: dynamics and control. Oxford: Oxford University Press, 1992.
- Fine PE. Herd immunity: history, theory, practice. Epidemiol Rev 1993;15:265–302.
- 107. Dietz K. The estimation of the basic reproduction number for infectious diseases. Stat Methods Med Res 1993;2:23–41.
- Coale AJ, Demeny P. Regional model life tables and stable populations. Princeton, NJ: Princeton University Press, 1966.
- United Nations. World population prospects: the 1994 revision. New York: United Nations, 1995. (ST/ESA/SER.S/145 and E.95.XIII.16).
- 110. Eichner M, Hadeler KP, Dietz K. Stochastic models for the eradication of poliomyelitis: minimum population size for

polio virus persistence. In: Isham V, Medley G, eds. Models for infectious human diseases: their structure and relation to data. Cambridge: Cambridge University Press, 1996.

- 111. Eichner M, Dietz K. Eradication of poliomyelitis: when can one be sure that polio virus transmission has been terminated? Am J Epidemiol 1996;143:816–22.
- 112. Patriarca PA, Sutter RW, Oostvogel PM. Outbreaks of paralytic poliomyelitis, 1976–1995. J Infect Dis 1997;175 (suppl 1):S165–72.
- 113. Nathanson N, Martin JR. The epidemiology of poliomyelitis: enigmas surrounding its appearance, epidemicity, and disappearance. Am J Epidemiol 1979;110:672–92.
- 114. Gard S, Bottiger M, Lagercrantz R. Vaccination with attenuated poliovirus type 1, the CHAT strain. In: Proceedings of the 1st International Conference on Live Poliovirus Vaccines. Washington, DC: Pan American Sanitary Bureau, 1959: 350-4.
- 115. Paul JR. The spread of attenuated poliovirus among household contacts. (Abstract). In: Poliomyelitis: papers and discussions presented at the 5th International Poliomyelitis Conference, Copenhagen, Denmark, July 26–28, 1960. Philadelphia: JB Lippincott Co, 1960:23–4.
- 116. Ghendon YZ, Sanakoyeva II. Comparison of the resistance of the intestinal tract to poliomyelitis virus (Sabin's strains) in persons after naturally and experimentally acquired immunity. Acta Virol 1961;5:265–73.
- 117. Schaap GJ, Bijkerk H, Coutinho RA, et al. The spread of wild poliovirus in the well-vaccinated Netherlands in connection with the 1978 epidemic. Prog Med Virol 1984;29: 124-40.
- 118. Poliomyelitis—United States, 1975–1984. MMWR Morb Mortal Wkly Rep 1986;35:180–2.
- 119. Isolation of wild poliovirus type 3 among members of a religious community objecting to vaccination—Alberta, Canada, 1993. MMWR Morb Mortal Wkly Rep 1993;42: 337–9.
- Chin KP, Lok AS, Wong LS, et al. Current seroepidemiology of hepatitis A in Hong Kong. J Med Virol 1991;34:191–3.
- Wu JS, Lu CF, Wu LZ, et al. Changing seroepidemiology of hepatitis A virus infection between two regions in Taiwan differing in socioeconomic status. J Formos Med Assoc 1993;92:812–15.
- 122. Kamel MA, Troonen H, Kapprell HP, et al. Seroepidemiology of hepatitis E virus in the Egyptian Nile delta. J Med Virol 1995;47:399–403.
- Swartz TA, Skalska P, Gerichter CG, et al. Routine administration of oral polio vaccine in a subtropical area. Factors possibly influencing sero-conversion rates. J Hyg Lond 1972;70:719-26.
- John TJ, Jayabal P. Oral polio vaccination of children in the tropics. I. The poor seroconversion rates and the absence of viral interference. Am J Epidemiol 1972;96:263–9.
- 125. Patriarca PA, Wright PF, John TJ. Factors affecting the immunogenicity of oral poliovirus vaccine in developing countries: review. Rev Infect Dis 1991;13:926–39.
- 126. Factors affecting the immunogenicity of oral poliovirus vaccine: a prospective evaluation in Brazil and The Gambia. J Infect Dis 1995;171:1097–106.
- 127. Osei Kwasi M, Afari EA, Mimura K, et al. Randomized, controlled trial of trivalent oral poliovirus vaccine (Sabin) starting at birth in Ghana. Bull World Health Organ 1995; 73:41-6.
- 128. Paul JR, Horstmann DM, Riordan JT, et al. The capacity of live attenuated polioviruses to cause human infection and to spread within families. In: Proceedings of the 2nd International Conference on Live Poliovirus Vaccines. Washington, DC: Pan American Sanitary Bureau, 1960: 174-84.
- 129. Hale JH, Lee LH, Gardner PS. A study of interference among enteroviruses during the mass immunization campaign with attenuated poliovirus type 2. In: Poliomyelitis: papers and discussions presented at the 5th International Poliomyelitis

Conference, Copenhagen, Denmark, July 26–28, 1960. Philadelphia: JB Lippincott Co, 1960:336–41.

- Voroshilova MK, Zhevandrova VI, Koroleva GA, et al. The effect of enterovirus carriage on results of oral immunization with live poliovirus vaccine from Sabin strains. In: Choumakov MP, ed. Oral live poliovirus vaccine. (Papers presented at the IVth Scientific Conference of the Institute of Poliomyelitis and Virus Encephalitis and the International Symposium on Live Poliovirus Vaccine, May 17-20, 1960). Moscow: Academy of Medical Sciences of the USSR, 1961:560-73.
   Drozdov SG, Shirman GA. Interaction of viruses in the
- 131. Drozdov SG, Shirman GA. Interaction of viruses in the intestinal tract of man. I. Interference between wild and vaccine poliovirus strains. Acta Virol 1961;5:210–19.
- 132. Przesmycki F, Dobrowolska H, Georgiades J, et al. Report on field trials with live attenuated poliomyelitis vaccine of Koprowski in Poland. Am J Hyg 1960;71:275–84.
- Ghendon Y, Robertson SE. Interrupting the transmission of wild polioviruses with vaccines: immunological considerations. Bull World Health Organ 1994;72:973–83.
- 134. Katz M, Plotkin SA. Minimal infective dose of attenuated poliovirus for man. Am J Public Health 1967;57:1837–40.
- 135. Verlinde JD, Wilterdink JB. A small-scale trial on vaccination and revaccination with live attenuated polioviruses in the Netherlands. In: Proceedings of the 1st International Conference on Live Poliovirus Vaccines. Washington, DC: Pan American Sanitary Bureau, 1959:355-66.
- 136. Lepow ML, Warren RJ, Gray N, et al. Effect of Sabin type 1 poliomyelitis vaccine administered by mouth to newborn infants. N Engl J Med 1961;264:1071–8.
- 137. Henry JL, Jaikaran ES, Davies JR, et al. A study of poliovaccination in infancy: excretion following challenge with live virus by children given killed or living poliovaccine. J Hyg (Cambridge) 1966;64:105-20.
- 138. Alexander JPJ, Gary HEJ, Pallansch MA. Duration of poliovirus excretion and its implications for acute flaccid paralysis surveillance: a review of the literature. J Infect Dis 1997;175(suppl 1):S176-82.
- 139. Melnick JL, Rennick V. Infectivity titers of enterovirus as found in human stools. J Med Virol 1980;5:205–20.
- 140. Zeitlenok NA, Lovtzevich EL, Bagdasaryan GA. Experimental study of the sensitivity to chlorine of attenuated and virulent poliomyelitis virus strains and their adsorption by various kinds of soil. In: Choumakov MP, ed. Oral live poliovirus vaccine. (Papers presented at the IVth Scientific Conference of the Institute of Poliomyelitis and Virus Encephalitis and the International Symposium on Live Poliovirus Vaccine, May 17–20, 1960). Moscow: Academy of Medical Sciences of the USSR, 1961:591–4.
- 141. Reichler MR, Patriarca PA. The place of outbreak response immunization in the control of poliovirus spread. Immunization strategies for poliomyelitis control: preliminary results of a review of the literature from 1900 to the present. (Working paper for the World Health Organization, June 7-11, 1993). Geneva, Switzerland: World Health Organization, 1993. (EPI/TECHCOMM/WP/93.5).
- 142. Peretz LG, Medvinskaya KG, Neznanskaya II. Interference between different strains of polioviruses in the light of general theory of microbial antagonism. In: Choumakov MP, ed. Oral live poliovirus vaccine. (Papers presented at the IVth Scientific Conference of the Institute of Poliomyelitis and Virus Encephalitis and the International Symposium on Live Poliovirus Vaccine, May 17-20, 1960). Moscow: Academy

of Medical Sciences of the USSR, 1961:580-3.

- 143. Drosdov SG, Shirman GA, Knyazeva TV. Interference between wild and vaccine strains of poliomyelitis virus in the human intestinal tract. In: Choumakov MP, ed. Oral live poliovirus vaccine. (Papers presented at the IVth Scientific Conference of the Institute of Poliomyelitis and Virus Encephalitis and the International Symposium on Live Poliovirus Vaccine, May 17-20, 1960). Moscow: Academy of Medical Sciences of the USSR, 1961:546-59.
- 144. Ramsay ME, Begg NT, Gandhi J, et al. Antibody response and viral excretion after live polio vaccine or a combined schedule of live and inactivated polio vaccines. Pediatr Infect Dis J 1994;13:1117-21.
- De Quadros CA, Andrus JK, Olive JM, et al. Polio eradication from the Western hemisphere. Annu Rev Public Health 1991;13:239-53.
- 146. Cossart YE. Evolution of poliovirus since introduction of attenuated vaccine. Br Med J 1977;1:1621-3.
- Sellwood J, Dadswell JV, Slade JS. Viruses in sewage as an indicator of their presence in the community. J Hyg Lond 1981;86:217-25.
- 148. Macdonald G. The epidemiology and control of malaria. London: Oxford University Press, 1957.
- World Health Organization. EPI information system. Global summary August, 1997. Geneva, Switzerland: World Health Organization, 1997. (WHO/EPI/GEN/97.02).
- Cohen Abbo A, Culley BS, Reed GW, et al. Seroresponse to trivalent oral poliovirus vaccine as a function of dosage interval. Pediatr Infect Dis J 1995;14:100-6.
- 151. Nishio O, Ishihara Y, Sakae K, et al. The trend of acquired immunity with live poliovirus vaccine and the effect of revaccination: follow-up of vaccinees for ten years. J Biol Stand 1984;12:1-10.
- 152. Onorato IM, Modlin JF, McBean AM, et al. Mucosal immunity induced by enhanced-potency inactivated and oral polio vaccines. J Infect Dis 1991;163:1-6.
- 153. Glezen WP, McCollough RH, Lamb GA, et al. Quantitative relationship of preexisting homotypic antibodies to excretion of poliovirus types 1, 2, and 3 following the feeding of trivalent attenuated poliovirus vaccine. Am J Epidemiol 1969;90: 146–56.
- Smith JW, Lee JA, Fletcher WB, et al. The response to oral poliovaccine in persons aged 16-18 years. J Hyg Lond 1976; 76:235-47.
- 155. Holguin AH, Reeves JS, Gelfand HM. Immunization of infants with the Sabin oral poliovirus vaccine. Am J Public Health 1962;52:600-10.
- 156. Bottiger M. A study of the sero-immunity that has protected the Swedish population against poliomyelitis for 25 years. Scand J Infect Dis 1987;19:595-601.
- 157. Black FL, Hierholzer WJ, Pinheiro F, et al. Evidence for persistence of infectious agents in isolated human populations. Am J Epidemiol 1974;100:230-50.
- 158. Parkman P. An assessment of the safety and efficacy implications of removing the type 2 strain from the trivalent oral poliovirus vaccine. Presented at the Stopping Polio Immunization Meeting of the World Health Organization, Geneva, Switzerland, March 31, 1997.
- 159. Cochi SL, Sutter RW, Kew OM, et al. A decision tree for stopping polio immunization. Technical consultation on the global eradication of poliomyelitis. Geneva, Switzerland: World Health Organization, 1997. (EPI/POLIO/TECH.97/WP.18).